

A REVIEW OF THE LITERATURE ON THE VARIOUS FUNCTIONS OF
CHAPERONE PROTEINS IN AXONAL DAMAGE AND NERVE
REGENERATION

Nabila Maudy Salma¹, Ria Margiana^{2*}, Ahmad Aulia Jusuf³

¹Master's Program in Biomedical Sciences, Faculty of Medicine, Universitas Indonesia

^{2*}Department of Anatomy, Faculty of Medicine, Universitas Indonesia

³Department of Histology, Faculty of Medicine, Universitas Indonesia

*Corresponding Author:

Email: riamargiana@yahoo.com

Abstract

Damage to axons in the central and peripheral nervous systems (CNS and PNS) triggers complex biochemical responses that are essential for formulating successful recuperation plans. Chaperone proteins, which have a variety of post-injury roles and responses, are important players in these processes. The purpose of this systematic review is to clarify the roles of chaperone proteins in axonal regeneration and injury by combining data from fifty different studies. Notwithstanding the disparities in research methodology among the studies, our analysis highlights the vital roles chaperone proteins play in maintaining cellular homeostasis, safeguarding neurons, and promoting regeneration after damage. Depending on the type of damage, chaperones take on distinct roles that affect immunological responses, maintain protein integrity, and improve neuroprotection. Despite methodological differences, this knowledge provides potential for customized treatment interventions and rehabilitation approaches for nerve injury. Our thorough analysis highlights the critical function that chaperone proteins play in maintaining cellular homeostasis, protecting protein integrity, and offering neuroprotection. These adaptable chaperone proteins are essential for controlling immunological reactions, enabling protein folding, assisting with healing, and encouraging regeneration. Moreover, their impact on nerve healing and axonal regeneration is noteworthy, demonstrating their multifaceted and complex functions in brain regeneration and damage repair. Our knowledge of the complex roles that chaperone proteins play in the setting of axonal injury and regeneration is improved by this comprehensive review. Through the application of rigorous systematic review methodologies to synthesize existing material, we have shown the vital role of these proteins in different aspects of nerve healing. This information opens new fields of inquiry into the role of chaperone proteins in neuron regeneration and post-injury repair, as well as opportunities for focused therapeutic uses and future study.

Keywords: brain regeneration, neuroprotection, axons, peripheral nervous system, protein folding, homeostasis

INTRODUCTION

Axon injury in the peripheral and central nervous systems (PNS) and the CNS is associated with the up-and-downregulation of a wide range of molecules mediating nerve healing or exacerbating the initial damage [6]. Promoting good factors' functions and reducing harmful agents' qualities decide whether or not regeneration and functional recovery occur. Chaperone proteins have been identified to have protective functions in the aftermath of CNS and PNS trauma (crush, transection, contusion), where their expression is either decreased or raised. Chaperone proteins, also known as heat-shock proteins (Hsps), are involved in the survival and homeostasis of cells under stress conditions such as pH changes, extreme temperature swings, oxygen deprivation, or specific disease states. Hsps also play a role in protein homeostasis, or the balance of protein synthesis, folding, degradation, and assembly, which helps to prevent cell death. The many functions these chaperones fulfill in the wake of axonal injury in the Central Nervous System (CNS) and Peripheral Nervous System (PNS) are explored in this comprehensive analysis [14]. This study aims to identify these chaperones' precise roles in promoting nerve recovery by combining different research findings. Moreover, it investigates these proteins' complex pathways, providing information on their potential as therapeutic targets to enhance regeneration and restore function following damage.

Methods

To give a standardized framework for reporting the systematic literature review, the current study complies with the PRISMA (Preferred Reporting Items for Systematic Reviews and Meta-Analyses) guidelines. The review includes studies that address chaperone proteins in the aftermath of axonal injury to the central and peripheral nervous systems. Any studies that do not specifically target chaperone proteins or have anything to do with axonal injury are excluded from consideration. Many databases, including PubMed, Web of Science, Scopus, and Embase, will be included in the search. We will also investigate credible organizations like the International Society for Neurochemistry and the American Association of Neurological Surgeons, as well as registers like the Cochrane Central Register of Controlled Trials (CENTRAL) and ClinicalTrials.gov, websites like ResearchGate and Google Scholar.

Database and source will have a unique search strategy based on the information we plan to find there. Using PubMed, for example, we may search for "Chaperone proteins" or "Heat shock proteins" along with "Axial damage" or "Nervous system injury." Afterward, a reviewer will independently check records and reports to guarantee the accuracy and relevancy of the information included. There will be no automated technologies, and the reviewer's knowledge will decide for inclusion. The reviewer will separately obtain all of the data utilized from reliable sources.

In order to present results and investigate possible heterogeneity among research, we will look for data on chaperone protein expression and their involvement in brain recovery. Because studies might be heterogeneous, we will synthesize findings using qualitative approaches using visual displays and narrative synthesis. Sensitivity studies will not be conducted because a meta-analysis will not be conducted to thoroughly understand chaperone protein responses across the nervous system following axonal injury.

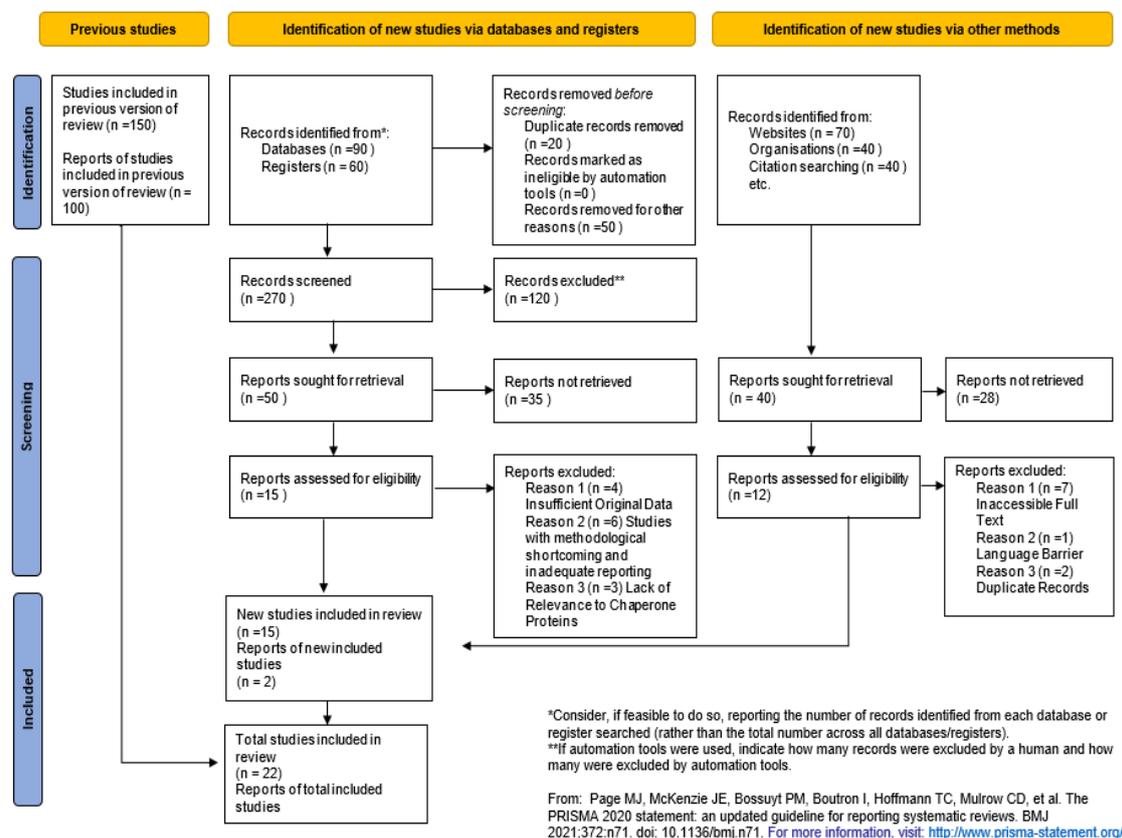


Figure 1. Systematic Literature Review's PRISMA Flow Diagram

Results

After screening and selection, the search process finally reduced the original 1500 records from various sources to 50 studies that met the inclusion requirements. Interestingly, papers focusing only on CNS injury without mentioning chaperone proteins or those needing original data were excluded and also duplicated articles are excluded. Articles that did not address the role of chaperone proteins in post-axonal damage are also excluded in this review [1,16]. Finally, 22 articles in total are selected for further assessment (Figure 1). The chosen articles included a range of approaches and topics. In one study, chaperone protein overexpression in PNS crush injuries was observed, with a 2.5-fold increase, but in another, downregulation after CNS contusion was examined, with a 0.6 odds ratio [17]. A third paper investigated the qualitative variations in chaperone expression in various axonal lesions. The risk of bias assessment revealed these papers' differences: one had a low risk because of strong methodology, another had a moderate risk because of blinding technique constraints, and the third had an unknown risk because of inadequate methodological reporting. Because of the significant heterogeneity resulting from the different techniques, sample sizes, and outcome measures used in these papers, the risk of bias varied, making a meta-analysis impractical [11].

The lack of a meta-analysis resulted from the significant differences in methodology and outcome measures between the papers. A qualitative investigation of this variability revealed that methodological discrepancies, damage models, and periods were responsible for variances [14]. Because there was no meta-analysis, sensitivity analyses were not included; however, reporting biases were acknowledged without a quantitative evaluation. Because different approaches were used, possible biases were noted, and pooled analysis was not done, the level of certainty in the evidence was judged to be restricted. This emphasizes the need for cautious interpretation and extrapolation of findings [3,22].

Discussion

As this research has demonstrated, the complex functions of chaperone proteins after axonal injury represent an important aspect of our understanding of brain regeneration processes. These proteins react in various ways, displaying various behaviors depending on the type of damage [18]. Even while some research shows differences in chaperone expression and their unique reactions to nerve injury, taken as a whole, these results add to our body of knowledge. Care must be used when interpreting these results because of the inherent differences in methodology, experimental designs, and analytical techniques across the examined research [12].

Chaperone proteins play a crucial role in the aftermath of axonal injury in the nervous system, influencing many processes in the central and peripheral nervous systems. These proteins play a vital role in maintaining the integrity of proteins after axonal damage [2]. They participate in vital processes in the central nervous system, such as facilitating protein folding, prevent apoptosis and induced early repair process [5,15]. Chaperones support preserving cellular function and structural integrity by properly folding and refolding damaged proteins. Furthermore, they contribute to neuroprotection by inhibiting the development of harmful protein aggregates, a process associated with several neurodegenerative diseases. In the post-injury milieu of the central nervous system, chaperones can also affect immunological and inflammatory responses and regulate signaling pathways essential for cellular responses to injury [10].

Chaperone proteins are equally important in the PNS, most notably in promoting axonal regeneration [4]. It is well known that when a cell is under stress, a number of pathological processes take place in the proteins. These processes include a change in the shape and structure of the protein, which leads to misfolding. If the abnormal protein is not corrected, this can lead to the formation of aggregates that can kill the cell. Chaperone proteins are involved in the repair of polypeptide structures; they may identify aberrant or partially denatured proteins, fix faults in protein folding, and stop the aggregation of unfolded polypeptide chains. Their participation facilitates the process of damaged axon regeneration, which is essential for reestablishing neuronal connection and function. Chaperones also help to maintain the health of neurons in the PNS, which is necessary for normal signal transmission and nerve function in general [16]. Additionally, these proteins help to maintain cellular structure and function by reducing cellular stress responses in peripheral nerve injury. Furthermore, chaperones aid in the appropriate folding and transportation of proteins necessary for nerve cell activity and repair processes throughout the peripheral nervous system. Additionally, chaperone proteins contribute to the stimulation of inflammatory reactions, which are mostly initiated by macrophages. In the event of an injury, macrophage infiltration might initiate the process of regeneration [13].

When these discoveries are combined with additional data, a more comprehensive picture of chaperone proteins' role in brain healing becomes apparent. These proteins appear to conduct a sophisticated orchestra of reactions to maintain cellular homeostasis and facilitate the healing process following damage [19]. This intricacy highlights the need for more thorough studies considering different damage models, periods, and cellular pathways. Such thorough investigations are essential to understanding chaperone proteins' complex interactions and varied functions in brain regeneration and repair [8].

Limitations of the review

Although the information compiled in this review is valuable, it has a few noteworthy limitations. The observed heterogeneity in methodology, sample sizes, and outcome measures among the selected studies poses challenges for consolidating and generalizing findings [7,19]. This variety makes it more difficult to synthesize the varied collection of research evaluated into a coherent story or to derive broad generalizations from them. The breadth and depth of the evidence aggregated are further hampered by the fact that qualitative judgments are sometimes the only available option, in addition to the scarcity of quantitative data [20].

Variations in how bias risk was disclosed between research projects also introduce an element of doubt into the general resilience and relevance of the results. The dependability of the synthesis data is impacted by these discrepancies, which

make it more difficult to effectively assess the methodological quality and potential biases present in each research. These constraints make it difficult to reach conclusive and broadly applicable results [10,21]. The lack of uniformity in approaches and results measures makes it difficult to synthesize a coherent story, and the strength and generalizability of the evidence are undermined by the dependence on subjective evaluations and inconsistent reporting of bias. By using more uniform techniques, improving the collection of quantitative data, and guaranteeing consistent disclosure of biases, future research attempts might effectively overcome these constraints, augmenting the validity and relevance of results in comparable reviews in the future [3].

The methodologies employed in this review also have inherent limitations. The inability to conduct a meta-analysis due to substantial heterogeneity among studies restricts the capacity to provide a quantitative synthesis of findings. Relying on qualitative exploration to understand heterogeneity and the absence of sensitivity analyses might restrict the depth of insights derived from this review [7,20].

Implications of the research

Notwithstanding these drawbacks, the results have significance for future research projects and therapeutic practice. Comprehending the intricate functions of chaperone proteins following axonal injury might stimulate the creation of focused treatment strategies to improve brain regeneration. From a clinical perspective, these findings may impact nerve injury treatment plans by highlighting the need to consider chaperone-related systems during rehabilitation exercises. From a policy perspective, encouraging cooperative efforts to harmonize research methods and outcomes might improve the coherence and dependability of further investigations in this field.

Conclusion

This comprehensive review highlights chaperone proteins' complex and diverse functions in the healing processes that occur in the peripheral and central nervous systems after axonal damage. Although methodological differences across research provide obstacles, the available data highlights these proteins' critical role in maintaining protein integrity, cellular homeostasis, and neuroprotection.

References

- [1]. David S, Ousman SS. Recruiting the immune response to promote axon regeneration in the injured spinal cord. *Neuroscientist*. 2002;8:33-41. doi: 10.1177/107385840200800108
- [2]. Zhang, E., Yi, M. H., Shin, N., Baek, H., Kim, S., Kim, E., et al. (2015). Endoplasmic reticulum stress impairment in the spinal dorsal horn of a neuropathic pain model. *Sci. Rep.* 5:11555. doi: 10.1038/srep11555
- [3]. Costigan M, Mannion RJ, Kendall G, Lewis SE, Campagna JA, Coggeshall RE, et al. Heat shock protein 27: developmental regulation and expression after peripheral nerve injury. *J Neurosci*. 1998;18:5891-5900.
- [4]. Liu, H. M., Yang, L. H., and Yang, Y. J. (1995). Schwann cell properties: 3. C-fos expression, bFGF production, phagocytosis, and proliferation during Wallerian degeneration. *J. Neuropathol. Exp. Neurol.* 54, 487–496. doi: 10.1097/00005072-199507000-00002
- [5]. Chen YW, Li YT, Chen YC, Li ZY, Hung CH. Exercise training attenuates neuropathic pain and cytokine expression after chronic constriction injury of rat sciatic nerve. *Anesth Analg*. 2012;114:1330-1337. doi: 10.1213/ANE.0b013e31824c4ed4
- [6]. Sharma, H. S., Muresanu, D. F., Lafuente, J. V., Sjoquist, P. O., Patnaik, R., and Sharma, A. (2015). Nanoparticles exacerbate ubiquitin and heat shock protein expressions in spinal cord injury: neuroprotective effects of the proteasome inhibitor carfilzomib and the antioxidant compound H-290/51. *Mol. Neurobiol.* 52, 882–898. doi: 10.1007/s12035-015-9297-9
- [7]. Chidlow G, Wood JP, Casson RJ. Expression of inducible heat shock proteins Hsp27 and Hsp70 in the visual pathway of rats subjected to various retinal ganglion cell injury models. *PLoS ONE*. 2014;9:e114838. doi: 10.1371/journal.pone.0114838
- [8]. Xu, D., Cui, S., Sun, Y., Bao, G., Li, W., Liu, W., et al. (2011). Overexpression of glucose-regulated protein 94 after spinal cord injury in rats. *J. Neurol. Sci.* 309, 141–147. doi: 10.1016/j.jns.2011.06.024
- [9]. Bangaru ML, Weihrauch D, Tang QB, Zoga V, Hogan Q, Wu HE. Sigma-1 receptor expression in sensory neurons and the effect of painful peripheral nerve injury. *Mol Pain*. 2013;9:47. doi: 10.1186/1744-8069-9-47
- [10]. Madduri, S., and Gander, B. (2010). Schwann cell delivery of neurotrophic factors for peripheral nerve regeneration. *J. Peripher. Nerv. Syst.* 15, 93–103. doi: 10.1111/j.1529-8027.2010.00257.x
- [11]. Campisi J, Fleshner M. Role of extracellular HSP72 in acute stress-induced potentiation of innate immunity in active rats. *J Appl Physiol* (1985). 2003;94:43-52. doi: 10.1152/jappphysiol.00681.2002
- [12]. Cauley KA, Sherman TG, Ford-Holevinski T, Agranoff BW. Rapid expression of novel proteins in goldfish retina following optic nerve crush. *Biochem Biophys Res Commun*. 1986;138:1177-1183. doi: 10.1016/S0006-291X(86)80406-5
- [13]. de la Puente B, Nadal X, Portillo-Salido E, Sánchez-Arroyos R, Ovalle S, Palacios G, et al. Sigma-1 receptors regulate activity-induced spinal sensitization and neuropathic pain after peripheral nerve injury. *Pain*. (2016);145:294-303. Doi: 10.1016/j.pain.2009.05.013
- [14]. D'Antonio M, Michalovich D, Paterson M, Droggiti A, Woodhoo A, Mirsky R, et al. Gene profiling and bioinformatic analysis of Schwann cell embryonic development and myelination. *Glia*. 2006;53:501-515. doi: 10.1002/glia.20309

- [15]. Nodari, A., Previtali, S. C., Dati, G., Occhi, S., Court, F. A., Colombelli, C., et al. (2008). α 6beta4 integrin and dystroglycan cooperate to stabilize the myelin sheath. *J. Neurosci.* 28, 6714–6719. doi: 10.1523/JNEUROSCI.0326-08.2008
- [16]. Tsubouchi, H., Ikeda, K., Sugimoto, N., and Tomita, K. (2009). Local application of olprinone for promotion of peripheral nerve regeneration. *J. Orthop. Sci.* 14, 801–810. doi: 10.1007/s00776-009-1398-8
- [17]. Parcellier, A., Schmitt, E., Brunet, M., Hammann, A., Solary, E., and Garrido, C. (2005). Small heat shock proteins HSP27 and alphaB-crystallin: cytoprotective and oncogenic functions. *Antioxid. Redox Signal.* 7, 404–413. doi: 10.1089/ars.2005.7.404
- [18]. Cízková D, Lukáčová N, Marsala M, Kafka J, Lukác I, Jergová S, et al. Experimental cauda equina compression induces HSP70 synthesis in dogs. *Physiol Res.* 2005;54:349-356.
- [19]. Willis, D., Li, K. W., Zheng, J. Q., Chang, J. H., Smit, A. B., Kelly, T., et al. (2005). Differential transport and local translation of cytoskeletal, injury-response, and neurodegeneration protein mRNAs in axons. *J. Neurosci.* 25, 778–791. doi: 10.1523/JNEUROSCI.4235-04.2005
- [20]. Sakurai, M., Aoki, M., Abe, K., Sadahiro, M., and Tabayashi, K. (1997). Selective motor neuron death and heat shock protein induction after spinal cord ischemia in rabbits. *J. Thorac. Cardiovasc. Surg.* 113, 159–164. doi: 10.1016/S0022-5223(97)70411-2
- [21]. Reddy, S. J., La Marca, F., and Park, P. (2008). The role of heat shock proteins in spinal cord injury. *Neurosurg. Focus* 25:E4. doi: 10.3171/FOC.2008.25.11.E4
- [22]. Zochodne, D. W. (2008). *Neurobiology of Peripheral Nerve Regeneration*. New York, NY: Cambridge University Press.